# 経済形質関連マーカーを用いた熊本系褐毛和種の遺伝的特性の解明

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# Elucidating genetic characteristics of the Kumamoto sub-breed of Japanese Brown cattle with DNA markers for economically important traits

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# ABSTRACT

The Kumamoto sub-breed of Japanese Brown cattle is famous for its heavy carcass weight, and increasing carcass weight is one of the most important breeding goals for this sub-breed. Genetic approaches may be especially useful for breeding this cattle and improving its traits. In this study, we elucidated the genetic characteristics of the Kumamoto sub-breed by analyzing the genotype and allele frequencies of DNA polymorphisms. Seven DNA polymorphisms were selected from the *SREBP1, SCD, FASN, EDG1, NCAPG, LYST,* and *F11* genes, which affect economically important traits. For genotyping, 112 offspring of 19 sires were collected. Our genotyping results show an extremely low allele frequency of the S allele of *SREBP1,* which affects fatty acid composition in intramuscular fat, in the Kumamoto sub-breed. Therefore, this polymorphism might not be useful for enhancing the breeding of this sub-breed. The causative mutation of *LYST* was not detected in our sampling population, suggesting a lower risk of Chediak-Higashi syndrome in the Kumamoto sub-breed. On the other hand, the Kumamoto sub-breed possessed high frequencies of desirable alleles of the *SCD* and *NCAPG* genes, which affect fatty acid composition and carcass weight, respectively, possibly because of artificial selection. Hence, these DNA markers could be used to improve economically important traits of the Kumamoto sub-breed. Our results also indicate that the causative mutation of factor XI deficiency might be common in the population because multiple sires were carriers of this disease. Since this disease may cause fetal losses, elimination of this disease from the Kumamoto sub-breed would be desirable.

Key words: economically important traits, genetic disease, Japanese Brown cattle, Kumamoto prefecture

# Introduction

Wagyu, a variety of beef cattle of Japanese origin, includes four genetically distinct breeds: Japanese Black, Japanese Brown, Japanese Shorthorn, and Japanese Polled (Gotoh *et al.* 2018). Japanese Brown cattle, raised mainly in the Kumamoto and Kochi prefectures, are the second most popular breed among the Wagyu breeds. The coat color patterns of the sub-breeds found in these two prefectures are apparently different, suggesting genetic isolation between

Correspondence: Hirokazu Matsumoto, Department of Animal Science, School of Agriculture, Tokai University, Toroku, Kumamoto, Toroku, Kumamoto 862-8652, Japan e-mail: yamato@tsc.u-tokai.ac.jp these two sub-breeds (Sasazaki *et al.* 2005; Matsumoto *et al.* 2020). The Kumamoto sub-breed is known to be easy to rear because of its calmer temperament and higher heat resistance (Ito *et al.* 2014; Imai *et al.* 2020). Moreover, weight gain is greater in this sub-breed, such that the average carcass weight of the Kumamoto sub-breed is heavier than that of Japanese Black cattle, which accounts for the majority of Wagyu breed cattle (Sasaki *et al.* 2006a). Genetic approaches may be useful for increasing carcass weight, which is an important breeding goal for the Kumamoto sub-breed, because economically important traits, including carcass weight, are highly heritable in this sub-breed (Sasaki *et al.* 2006a; Sumio 2007). However, information on their genetic characteristics is limited.

Various DNA polymorphisms that influence economically important traits in cattle have been identified. and some of them are utilized as DNA markers for markerassisted selection. Taniguchi et al. (2004) identified a missense variant in the SCD (stearoyl-CoA desaturase) gene, which affects fatty acid composition of intramuscular fat of Japanese Black cattle. In this variant, the 293rd amino acid changes from Val to Ala. This gene influences the mono-unsaturated fatty acid (MUFA) content that contributes to the tenderness and flavor of the meat, which is regarded as one of the most economically important traits (Melton et al. 1982). The Ala allele of the SCD gene increases MUFA percentage in a dose-dependent manner. Furthermore, this single nucleotide polymorphism (SNP) does not negatively affect other economically important traits (Ohsaki et al. 2009). Subsequent studies using various beef cattle breeds have confirmed the effects of this SNP on fatty acid composition (Orrù et al. 2011; Li et al. 2012).

Because most economically important traits are quantitative traits controlled by various genes, development of additional DNA markers could help beef cattle breeding. Hence, many genes affecting economically important traits have been identified. For example, *fatty acid synthase* (*FASN*) and *sterol regulatory element binding protein 1* (*SREBP1*) are other genes associated with fatty acid composition (Hoashi *et al.* 2007; Abe *et al.* 2009). Similarly, the beef marbling standard (BMS), one of the most economically important traits, is influenced by the *sphingosine-1-phosphate receptor 1 (EDG1)* gene (Yamada *et al.* 2009). The *non-SMC condensin-I complex, subunit G* (*NCAPG*) gene influences carcass weight (Setoguchi *et al.*  2009). In Wagyu populations, several genetic diseases, which may have negative impacts on beef cattle production, and their causative genes have been reported. These include *lysosomal trafficking regulator (LYST)* gene, which is the causative gene for Chediak-Higashi syndrome and *coagulation factor XI (F11)* gene, which is the causative gene for factor XI deficiency (Kunieda *et al.* 1999, Kunieda *et al.* 2005). The clinical symptoms of these diseases are relatively mild, suggesting that carriers of the causative mutations might be found in the population of the Kumamoto sub-breed of Japanese Brown cattle. Investigation of mutations in these genes will be useful for breeding the Kumamoto sub-breed.

In this study, to elucidate the genetic characteristics of the Kumamoto sub-breed of Japanese Brown cattle, we analyzed the genotype and allele frequencies of variants in seven genes: *SREBP1* g.101\_185ins (rs133958066), *SCD* c.878T>C: p.Val293Ala (rs41255693), *FASN* c.5854A>G: p.Thr1952Ala (rs208645216), *EDG1* c.-312A>G (rs211101524), *NCAPG* c.1326T>G: p.Ile442Met (rs109570900), *LYST* c.6044A>G: p.His2015Arg (AB026620), and *F11* c.870\_884ins (AB196307). Subsequently, we performed a chi-square test for deviation from Hardy-Weinberg equilibrium (HWE) to reveal the effects of each variant in this sub-breed. Our results might help determine an effective breeding strategy for this subbreed.

## Materials and Methods

#### Animals

In this study, 112 cattle (84 steers and 28 heifers), all of which belong to the Kumamoto sub-breeds of Japanese Brown cattle, were analyzed. Their meat was commercially purchased from Toyozumi shokuniku, a meat store in Kumamoto Prefecture, in 2018 and 2019. Genomic DNA samples were extracted using the standard phenolchloroform method for genotyping. These cattle were derived from 19 sires, and 56 of them were offspring of a single sire (sire A). Hence, the results of genotyping were shown as 112 cattle (sampling population), offspring of 18 sires (Group I) and offspring of sire A (Group II). The genotyping results of Group I were compared with other breeds because these cattle were assumed to reflect the characteristics of Kumamoto sub-breed. The genotyping results of Group II were also shown because the genetic characteristics of sire A may provide important information for breeding improvement of Kumamoto sub-breed.

## Genotyping

Genotyping for the nucleotide substitutions in SCD, FASN, EDG1, NCAPG, and LYST was performed by the restriction fragment length polymorphism method. To detect the SREBP1 and F11 polymorphisms, PCR was carried out. We genotyped SREBP1, SCD, FASN, EDG1, LYST and F11 genes according to Matsuhashi et al. (2011), Ookura et al. (2013), Sigintuya et al. (2014), Kaneda et al. (2011), Kunieda et al. (1999), and Kunieda et al. (2005), respectively. We designed the primer set to amplify the region including the NCAPG SNP based on the reference sequence (GenBank NC 037333.1) in Oligo7 (Molecular Biology Insights, Vondelpark, CO). Go-Tag® (Promega Corporation, Madison, WI) was used as the PCR enzyme and PCR was performed with the following conditions: 35 cycles at 95 °C for 30 sec, annealing temperatures given in Table 1 for 30 sec, and 72 °C for 30 sec. To amplify the region including EDG1 c.-312A>G, an extension procedure was performed for 60 sec. For F11 genotyping, the extension procedure was prolonged to 70 sec. Subsequent restriction enzyme reaction was performed at 37 °C for 15 min. All restriction enzymes were purchased from New England Biolabs (Ipswich, MA, USA). Primer sequences and restriction enzymes are listed in Table 1. After genotyping, the chi-square test ( $\chi^2$ ) was used to test deviation from HWE.

## **Results and Discussion**

We carried out genotyping for seven genes, but two of them were not polymorphic in the sampling population (Table 2). One of them was SREBP1, g.101 185ins, which has an 84-bp in/del polymorphism in intron 5 (Hoashi et al. 2007). SREBP1 is a transcription factor involved in fatty acid synthesis (Shimano 2001), and this polymorphism is thought to affect fatty acid composition. Hoashi et al. (2007) reported that the S allele (Short type) positively affects fatty acid composition. However, Ookura et al. (2013) reported that the L allele (long type) has a positive effect. The effect of SREBP1 g.101 185ins on the Kumamoto sub-breed is unknown, but this in/del cannot be used for breeding, because most individuals in the Kumamoto sub-breed were assumed to possess the L allele. On the other hand, the S allele frequency was 0.42 in the Kochi sub-breed (Kaneda et al. 2011), suggesting that these sub-breeds have different breeding histories. The other SNP that we detected was LYST c.6044A>G. This SNP is a missense mutation (p. His2015Arg) in exon 18, and the Arg allele is the causative allele of Chediak-Higashi syndrome, an autosomal recessive bleeding disorder (Kunieda et al. 1999). This genetic disease was found in several breeds, including Japanese Black cattle. However, we did not detect any individuals with an Arg allele from this population, suggesting that the risk of this genetic disease might be low in the Kumamoto subbreed.

The SCD enzyme converts saturated fatty acids into MUFAs in adipocytes (Kim and Ntambi 1999), and the Ala allele of p.Val293Ala has a positive effect on MUFA content (Taniguchi *et al.* 2004). Allele frequency of the Ala allele

| Table | 1. | Primer se | equences | and | restriction | enzymes t | for g | genotyping |  |
|-------|----|-----------|----------|-----|-------------|-----------|-------|------------|--|
|       |    |           |          |     |             |           | ,     |            |  |

| SNP          | Sequence $(5' \rightarrow 3')$                         | Tm (°C)* | Enzyme | Size (bp)       |
|--------------|--|----------|--------|-----------------|
| SREBP1       | F: CCACAACGCCATCGAGAAACGCTAC                           | 15       |        | L: 432          |
| g.101_185ins | R: GGCCTTCCCTGACCACCCAACTTAG                           | 65       | -      | S: 348          |
| SCD          | F: GTGGAAAATCAGGTAGGTCTCAGCG                           | (0)      | 4 - :T | T: 237, 370     |
| c.878T>C     | R: CAAGCAAGACTACCACCCAGATCAG                           | 60       | ACII   | C: 237, 58, 312 |
| FASN         | F: CTCACTGTCTGTCCCACAGG                                | (0)      | III.al | A: 243          |
| c.5854A>G    | R: GCCTTTGGAGGGCTTCTTAG                                | 60       | rinal  | G: 94, 149      |
| EDG1         | F: AAAGAAGCCACTCAGCCTCAGCG                             | 62       | Maal   | A: 77, 348      |
| c312A>G      | R: GTGAGCCCAGAACAATCCGATCC                             | 62       | MSCI   | G: 425          |
| NCAPG        | F: ACTCTACCTACCACACCAATATCCCT <u>C</u> AT <sup>†</sup> | 60       | Malli  | T: 177          |
| c.1326T>G    | R: AAGCCAAAAGCACTGAAAACA                               | 00       | mann   | G: 30, 147      |
| LYST         | F: GAAAATTACAGCAGAAGTCCTTGG                            | 60       | Fold   | A: 66, 42       |
| c.6044A>G    | R: TGACAAACATAAGTATTAGTAGGAGG                          | 60       | I'OKI  | G: 108          |
| F11          | F: TCACATCTCAATATGTGCTTCTGC                            | 55       |        | L: 110          |
| c.870_884ins | R: TCTACGATGTCCAGTTCTTCTCC                             | 22       | -      | S: 95           |

<sup>†</sup>Underline indicates a mismatch nucleotide to introduce the NlaIII recognition site into the PCR product.

\* Annealing temperature for the PCR reaction.

has been previously estimated in various cattle breeds: 0.76 in Japanese Black cattle, 0.83 in Angus cattle, and 0.49 in Simmental cattle (Kaneda et al. 2011; Orrù et al. 2011; Nishimaki et al. 2016). The frequencies of this allele in the Kumamoto sub-breed were 0.81 in the sampling population, 0.74 in Group I, and 0.89 in Group II (Table 2, 3). The high frequency of the Ala allele of 0.81 in the sampling population was contributed by Group II: offspring of sire A constituted half of our sample population, and 77% of them were homozygous for the Ala allele. Moreover, Group I showed deviation from the HWE (Table 3). Our data indicated that the genotype of sire A might be homozygous for the Ala allele and other sires might have more than one Ala allele. SCD c.878T>C (p.Val293Ala) has been reported to improve MUFA content in Japanese Black, Fleckvieh, Hanwoo, and Holstein cattle (Barton et al. 2010: Oh et al. 2011; Mannen 2012). Therefore, this SNP may also have a desirable effect on MUFA content of the Kumamoto sub-breed, and this effect may be the cause of the deviation from the HWE. However, the correlation between the polymorphism and fatty acid composition in the Kumamoto sub-breed remains to be elucidated.

The Ala allele of *FASN* p.Thr1952Ala, a missense mutation in exon 34, has been reported to have a positive effect on fatty acid composition (Abe *et al.* 2009). Frequency of this allele was 0.47 in Japanese Black cattle, 0.98 in Angus cattle, and 0.92 in Hereford cattle (Kaneda *et al.* 2011). The frequencies of this allele in the Kumamoto sub-breed were 0.43 in sampling population, 0.41 in Group I, and 0.46 in Group II (Table 2, 3), suggesting that this SNP

might improve the fatty acid composition of this sub-breed. However, further studies are required to fully understand the effect of this SNP in the Kumamoto sub-breed.

Expression of the *EDG1* gene, which is involved in blood vessel formation (Liu *et al.* 2000), is higher in highmarbled cattle than in low-marbled cattle (Sasaki *et al.* 2006b). An SNP in the untranslated region, c.-312A>G, might control *EDG1* expression, and cattle with the G allele show higher BMS (Yamada *et al.* 2009). The frequencies of this allele in the Kumamoto sub-breed were 0.05 in sampling population, 0.06 in Group I, and 0.05 in Group II (Table 2, 3). Our genotyping revealed that the G allele was quite rare in the Kumamoto sub-breed of Japanese Brown cattle. The frequency of this allele in other breeds, such as Angus and Hereford cattle, was also low (Kaneda *et al.* 2011). In contrast, Japanese Black cattle, known as highmarbled cattle, showed a higher G allele frequency of 0.41 (Watanabe *et al.* 2009).

The *NCAPG* gene encodes a condensin subunit that plays an important role in cell growth (Dej *et al.* 2004). Cattle with the Met allele of p.Ile442Met, a missense mutation in exon 9, show heavier carcass weight (Setoguchi *et al.* 2009). The frequencies of the Met allele in the Kumamoto sub-breed were 0.60 in sampling population, 0.43 in Group I, and 0.76 in Group II (Table 2, 3). The frequencies of this allele in the other breeds were as follows: 0.25 in Japanese Black cattle, 0.40 in Angus cattle, and 0.67 in Simmental cattle (Glenske *et al.* 2011; Nishimaki *et al.* 2016). This difference in the allele frequency might explain why the carcass weight of Japanese Brown and Simmental

| Table 2. Genotype and allele frequency in Kumamoto sub | <ul> <li>breed of Japanese Brown cattle</li> </ul> |
|--|--|
|--|--|

| CNID         | Sampling population (n = 112) |                    |                    |      |      |  |  |  |  |
|--------------|-------------------------------|--------------------|--------------------|------|------|--|--|--|--|
| SNP -        |                               | Allele fi          | Allele frequency   |      |      |  |  |  |  |
|              | S/S (n = 0)                   | S/L (n = 0)        | L/L (n = 112)      | S    | L    |  |  |  |  |
| SKEDF1 -     | 0.00                          | 0.00               | 1.00               | 0.00 | 1.00 |  |  |  |  |
| SCD          | Ala/Ala $(n = 70)$            | Ala/Val $(n = 42)$ | Val/Val $(n = 0)$  | Ala  | Val  |  |  |  |  |
| SCD -        | 0.63                          | 0.37               | 0.00               | 0.81 | 0.19 |  |  |  |  |
| EASN         | Ala/Ala $(n = 23)$            | Ala/Thr $(n = 50)$ | Thr/Thr $(n = 39)$ | Ala  | Thr  |  |  |  |  |
| FASIV -      | 0.21                          | 0.45               | 0.34               | 0.43 | 0.57 |  |  |  |  |
| EDCI         | G/G (n = 0)                   | G/A(n = 11)        | A/A (n = 101)      | G    | А    |  |  |  |  |
| EDGI -       | 0.00                          | 0.1                | 0.9                | 0.05 | 0.95 |  |  |  |  |
| NCADO        | Met/Met $(n = 42)$            | Met/Ile $(n = 49)$ | Ile/Ile $(n = 21)$ | Met  | Ile  |  |  |  |  |
| NCAPG -      | 0.38                          | 0.44               | 0.18               | 0.6  | 0.4  |  |  |  |  |
| IVET         | His/His (n = 112)             | His/Arg $(n = 0)$  | Arg/Arg (n = 0)    | His  | Arg  |  |  |  |  |
| LISI -       | 1.00                          | 0.00               | 0.00               | 1.00 | 0.00 |  |  |  |  |
| E11          | S/S (n = 55)                  | S/L (n = 50)       | L/L (n = 7)        | S    | L    |  |  |  |  |
| <i>F11</i> – | 0.49                          | 0.45               | 0.06               | 0.71 | 0.29 |  |  |  |  |

| SNP   |                    | Gr              | oup I (n = 56 | )         |        | Group II $(n = 56)$ |                    |          |          |        |        |  |
|-------|--------------------|-----------------|---------------|-----------|--------|---------------------|--------------------|----------|----------|--------|--------|--|
|       | C                  |                 |               |           | Allele |                     | Genotype frequency |          |          |        | Allele |  |
|       | Genotype frequency |                 |               | frequency |        |                     |                    |          |          | f      |        |  |
|       | Ala/Ala            | Ala/Val         | Val/Val       | Ala       | Val    |                     | Ala/Ala            | Ala/Val  | Val/Val  | . A la | Val    |  |
| SCD   | (n = 27)           | (n = 29)        | (n = 0)       | Ala Val   | vai    | 6.84**              | (n = 43)           | (n = 13) | (n = 0)  | Ala    | vai    |  |
|       | 0.48               | 0.52            | 0.00          | 0.74      | 0.26   |                     | 0.77               | 0.23     | 0.00     | 0.89   | 0.11   |  |
| FASN  | Ala/Ala            | Ala/Thr         | Thr/Thr       | Ala       | Th-    | 0.28                | Ala/Ala            | Ala/Thr  | Thr/Thr  | Ala    | Thr    |  |
|       | (n = 10)           | (n = 25)        | (n = 21)      | Ala       | 1 111  |                     | (n = 13)           | (n = 25) | (n = 18) |        |        |  |
|       | 0.18               | 0.45            | 0.37          | 0.41      | 0.59   | _                   | 0.23               | 0.45     | 0.32     | 0.46   | 0.54   |  |
| EDG1  | G/G                | G/A             | A/A           | C         | ٨      | 0.18                | G/G                | G/A      | A/A      | G      | А      |  |
|       | (n = 0)            | (n = 6)         | (n = 50)      | U         | A      |                     | (n = 0)            | (n = 5)  | (n = 51) |        |        |  |
|       | 0.00               | 0.11            | 0.89          | 0.06      | 0.94   |                     | 0                  | 0.09     | 0.91     | 0.05   | 0.95   |  |
| NCAPG | Met/Met            | Met/Met Met/Ile |               | Mat       | П.     |                     | Met/Met            | Met/Ile  | Ile/Ile  | Mat    | 11-    |  |
|       | (n = 10)           | (n = 28)        | (n = 18)      | wiet      | ne     | 0.02                | (n = 32)           | (n = 21) | (n = 3)  | Wiet   | ne     |  |
|       | 0.18               | 0.5             | 0.32          | 0.43      | 0.57   |                     | 0.57               | 0.38     | 0.05     | 0.76   | 0.24   |  |
| F11   | S/S                | S/L             | L/L           | S         | т      | 0.03                | S/S                | S/L      | L/L      | S      | L      |  |
|       | (n = 31)           | (n = 21)        | (n = 4)       | 5         | L      |                     | (n = 24)           | (n = 29) | (n = 3)  |        |        |  |
|       | 0.55               | 0.38            | 0.07          | 0.74      | 0.26   |                     | 0.43               | 0.52     | 0.05     | 0.69   | 0.31   |  |

Table 3. Genetic characteristics of offspring of 18 sires (Group I) and offspring of sire A (Group II)

\* Chi-square test (  $\chi$  2) with one degree of freedom (df = 1) was used to test deviation from HWE \*\* significant at p < 0.05 (  $\chi$  2 > 3.84, df = 1)

cattle is heavier than that of Japanese Black and Angus cattle (Sasaki *et al.* 2006a; Kause *et al.* 2015).

Factor XI deficiency is an autosomal recessive bleeding disorder. The causative mutation of this genetic disease is F11 c.870 884ins, a 15-bp insertion in exon 9 (Kunieda et al. 2005). The phenotype of this disease is mild, and individuals with the causative L allele (insertion type) grow normally in most cases; hence, this mutation tends to escape artificial selection (Ohba et al. 2008). While the L allele frequency was 0.38 in Japanese Black cattle (Ookura et al. 2013), the frequencies of 0.29 in sampling population, 0.26 in Group I, and 0.31 in Group II were estimated in the Kumamoto sub-breed in the current study. This suggests that multiple sires, including sire A, were carriers of factor XI deficiency (Table 3). Although this disease does not influence carcass traits (Ookura et al. 2013), factor XI deficiency could have negative economic effects because of the association between fetal losses and the L allele (Ogata et al. 2014). Therefore, elimination of this allele from Japanese Brown cattle is desirable.

In conclusion, we analyzed the genotype and allele frequencies of seven polymorphisms in the Kumamoto subbreed of Japanese Brown cattle in the present study. Our sample population showed polymorphisms in *SCD*, *FASN*, *EDG1*, *NCAPG*, and *F11*. The polymorphisms of *SCD*, *FASN*, and *NCAPG* might be useful for breeding the Kumamoto sub-breed. The deviation from HWE in *SCD* suggests the necessity to elucidate the effect of this polymorphism on fatty acid composition of the Kumamoto sub-breed. The heavy carcass weight of the Kumamoto sub-breed may be due to the high allele frequency of the desirable *NCAPG* allele. In addition, the mutant allele causing factor XI deficiency was found to be widespread in the Kumamoto sub-breed.

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## References

Abe T, Saburi J, Hasebe H, Nakagawa T, Misumi S, Nade T, Nakajima H, Shoji N, Kobayashi M, Kobayashi E.
2009. Novel mutations of the *FASN* gene and their effect on fatty acid composition in Japanese Black beef. Biochemical Genetics, 47: 397-411.

- Barton L, Kott T, Bures D, Rehák D, Zahrádková R, Kottová B. 2010. The polymorphisms of stearoyl-CoA desaturase (SCD1) and sterol regulatory element binding protein-1 (SREBP-1) genes and their association with the fatty acid profile of muscle and subcutaneous fat in Fleckvieh bulls. Meat Science, 85: 15-20.
- Dej KJ, Ahn C, Orr-Weaver TL. 2004. Mutations in the Drosophila condensin subunit dCAP-G: defining the role of condensin for chromosome condensation in mitosis and gene expression in interphase. Genetics, 168: 895-906.
- Glenske K, Brandt H, Erhardt G. 2011. Association of the Ile-442-Met substitution in NCAPG with birth weight in German Angus and German Simmental cattle (Brief Report). Archives Animal Breeding, 54: 104-106.
- Gotoh T, Nishimura T, Kuchida K, Mannen H. 2018. The Japanese Wagyu beef industry: current situation and future prospects - A review. Asian-Australasian Journal of Animal Sciences, 31: 933-950.
- Hoashi S, Ashida N, Ohsaki H, Utsugi T, Sasazaki S, Taniguchi M, Oyama K, Mukai F, Mannen H. 2007. Genotype of bovine sterol regulatory element binding protein-1 (SREBP-1) is associated with fatty acid composition in Japanese Black cattle. Mammalian Genome, 18: 880-886.
- Imai S, Sekiguchi R, Matsumoto K, Kashimura A, Inenaga T, Moritomo Y. 2020. Behavioral analysis handling and response to human in Japanese Black and Brown cattle at the livestock market. Proceedings of School of Agriculture Tokai University, 39: 21-26.
- Ito S, Iwashita R, Hagiwara S, Ymamoto N, Sakumoto R, Tani M, Okamoto C, Yayou K. 2014. Comparison of the maintenance behaviours of Japanese Brown and Japanese Black cows in pastures during the summer. Animal Behaviour and Management, 50: 162-168.
- Kaneda M, Lin BZ, Sasazaki S, Oyama K, Mannen H. 2011. Allele frequencies of gene polymorphisms related to economic trait in Bos taurus and Bos indicus cattle breeds. Animal Science Journal, 82: 717-721.
- Kause A, Mikkola L, Strandén I, Sirkko K. 2015. Genetic parameters for carcass weight, conformation and fat in five beef cattle breeds. Animal, 9: 35-42.
- Kim YC, Ntambi JM. 1999. Regulation of *stearoyl-CoA* desaturase genes: role in cellular metabolism and

preadipocyte differentiation. Biochemical and Biophysical Research Communications, 266: 1-4.

- Kunieda T, Nakagiri M, Takami M, Ide H, Ogawa H. 1999. Cloning of bovine LYST gene and identification of a missense mutation associated with Chediak-Higashi syndrome of cattle. Mammalian Genome, 10: 1146-1149.
- Kunieda M, Tsuji T, Abbasi AR, Khalaj M, Ikeda M, Miyadera K, Ogawa H, Kunieda T. 2005. An insertion mutation of the bovine *F11* gene is responsible for factor XI deficiency in Japanese black cattle. Mammalian Genome, 16: 383-389.
- Li C, Aldai N, Vinsky M, Dugan ME, McAllister TA. 2012. Association analyses of single nucleotide polymorphisms in bovine *stearoyl-CoA desaturase and fatty acid synthase* genes with fatty acid composition in commercial cross-bred beef steers. Animal Genetics, 43: 93-97.
- Liu Y, Wada R, Yamashita T, Mi Y, Deng CX, Hobson JP, Rosenfeldt HM, Nava VE, Chae SS, Lee MJ, Liu CH, Hla T, Spiegel S, Proia RL. 2000. Edg - 1, the G protein - coupled receptor for sphingosine - 1 phosphate, is essential for vascular maturation. Journal of Clinical Investigation, 106: 951-961.
- Mannen H. 2012. Genes associated with fatty acid composition of beef. Food Science and Technology Research, 18: 1-6.
- Matsuhashi T, Maruyama S, Uemoto Y, Kobayashi N, Mannen H, Abe T, Sakaguchi S, Kobayashi E. 2011.
  Effects of bovine *fatty acid synthase, stearoyl-coenzyme* A desaturase, sterol regulatory element-binding protein 1, and growth hormone gene polymorphisms on fatty acid composition and carcass traits in Japanese Black cattle. Journal of Animal Science, 89: 12-22.
- Matsumoto H, Kojya M, Takamuku H, Kimura S, Atsushi K, Imai S, Yamauchi K, Ito S. 2020. *MC1R* c.310G>- and c.871G > A determine the coat color of Kumamoto subbreed of Japanese Brown cattle. Animal Science Journal, 91: e13367.
- Melton SL, Amiri M, Davis GW, Backus WR. 1982. Flavor and chemical characteristics of ground beef from grass-, forage-, grain- and grain-finished steers. Journal of Animal Science, 55: 77-87.
- Nishimaki T, Ibi T, Siqintuya, Kobayashi N, Matsuhashi T, Akiyama T, Yoshida E, Imai K, Matsui M, Uemura K,

Eto H, Watanabe N, Fujita T, Saito Y, Komatsu T, Hoshiba H, Mannen H, Sasazaki S, Kunieda T. 2016. Allelic frequencies and association with carcass traits of six genes in local subpopulations of Japanese Black cattle. Animal Science Journal, 87: 469-476.

- Ogata Y, Siqintuya, Moritomo Y, Ohba Y, Kunieda T, Kitagawa H. 2014. Relationship between fetal losses and intrinsic coagulation disorder in Japanese Black cattle. Journal of the Japan Veterinary Medical Association, 67: 54-58.
- Oh DY, Lee YS, La BM, Yeo JS. 2011. Identification of the SNP (single nucleotide polymorphism) of the *stearoyl-CoA desaturase (SCD)* associated with unsaturated fatty acid in Hanwoo. Asian-Australasian Journal of Animal Sciences, 24: 757-765.
- Ohba Y, Takasu M, Nishii N, Takeda E, Maeda S, Kunieda T, Kitagawa H. 2008. Pedigree analysis of factor XI deficiency in Japanese black cattle. Journal of Veterinary Medical Science, 70: 297-299.
- Ohsaki H, Tanaka A, Hoashi S, Sasazaki S, Oyama K, Taniguchi M, Mukai F, Mannen H. 2009. Effect of *SCD* and *SREBP* genotypes on fatty acid composition in adipose tissue of Japanese Black cattle herds. Animal Science Journal, 80: 225-232.
- Ookura K, Akiyama T, Yoshida E, Fukushima M, Iwamoto E, Oka A, Matsumoto H, Sasazaki S, Oyama K, Mannen H. 2013. Effects of genes on economically important traits of Japanese Black cattle in Hyogo population. Nihon Chikusan Gakkaiho, 84: 157-162.
- Orrù L, Cifuni GF, Piasentier E, Corazzin M, Bovolenta S, Moioli B. 2011. Association analyses of single nucleotide polymorphisms in the *LEP* and *SCD1* genes on the fatty acid profile of muscle fat in Simmental bulls. Meat Science, 87: 344-348.
- Sasaki Y, Miyake T, Gaillard C, Oguni T, Matsumoto M, Ito M, Kurahara T, Sasae Y, Fujinaka K, Ohtagaki S, Dougo T. 2006a. Comparison of genetic gains per year for carcass traits among breeding programs in the Japanese Brown and the Japanese Black cattle. Journal of Animal Science, 84: 317-323.
- Sasaki Y, Nagai K, Nagata Y, Doronbekov K, Nishimura S, Yoshioka S, Fujita T, ShigaK, Miyake T, TaniguchiY, Yamada T. 2006b. Exploration of genes showing intramuscular fat deposition-associated expression changes in musculus longissimus muscle. Animal

Genetics, 37: 40-46.

- Sasazaki S, Usui M, Mannen H, Hiura C, Tsuji S. 2005. Allele frequencies of the extension locus encoding the melanocortin - 1 receptor in Japanese and Korean cattle. Animal Science Journal, 76: 129-132.
- Setoguchi K, Furuta M, Hirano T, Nagao T, Watanabe T, Sugimoto Y, Takasuga A. 2009. Cross-breed comparisons identified a critical 591-kb region for bovine carcass weight QTL (*CW-2*) on chromosome 6 and the Ile-442-Met substitution in *NCAPG* as a positional candidate. BMC Genetics, 10: 43.
- Shimano H. 2001. Sterol regulatory element-binding proteins (SREBPs): transcriptional regulators of lipid synthetic genes. Progress in Lipid Research, 40: 439-452.
- Siqintuya, Nishimaki T, Ibi T, Tsuji T, Yoneda K, Oshima I, Katahira K, Mannen H, Shimogiri T, Kanemaki M, Kunieda T. 2014. Allelic distributions of genes involved in economical traits, hereditary disorder, and coat color in a population of Kuchinoshima cattle. The Journal of Animal Genetics, 42: 11-19.
- Sumio Y. 2007. Improvement and present state of Japanese Brown cattle and prospect in the future. The Journal of Animal Genetics, 35: 141-146.
- Taniguchi M, Utsugi T, Oyama K, Mannen H, Kobayashi M, Tanabe Y, Ogino A, Tsuji S. 2004. Genotype of *stearoyl-coA desaturase* is associated with fatty acid composition in Japanese Black cattle. Mammalian Genome, 15: 142-148.
- Watanabe N, Yoshioka S, Itoh M, Satoh Y, Furuta M, Komatsu S, Sumio Y, Fujita T, Yamada T, Sasaki Y. 2009. The G allele at the c.-312A>G SNP in the *EDG1* gene associated with high marbling in Japanese Black cattle is at a low frequency in breeds not selected for marbling. Animal Genetics, 40: 579.
- Yamada T, Itoh M, Nishimura S, Taniguchi Y, Miyake T, Sasaki S, Yoshioka S, Fujita T, Shiga K, Morita M, Sasaki Y. 2009. Association of single nucleotide polymorphisms in the *endothelial differentiation sphingolipid G-protein-coupled receptor 1* gene with marbling in Japanese Black beef cattle. Animal Genetics, 40: 209-216.